
Structure and organization of two linked ribosomal protein genes in yeast

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ABSTRACT

The genes encoding yeast ribosomal proteins rp28 and S16A are linked and occur duplicated in the yeast genome. In both gene pairs the genes are approximately 600 bp apart and are both transcribed in the same direction. Both ribosomal protein genes resemble other ribosomal protein genes studied so far in many structural aspects. The genes are interrupted by an intron near the 5'-end of their coding sequence. In addition the flanking regions contain several conserved sequence elements, which may function in transcription initiation and termination.

In agreement with findings concerning other cloned yeast ribosomal protein genes, upstream homology blocks occur that may be involved in coordinate control of ribosomal protein gene transcription. The complete pattern of conserved and diverged sequences between the two duplicate gene pairs is presented.

INTRODUCTION

Yeast ribosomes are composed of 4 ribosomal RNAs and about 75 ribosomal proteins. For an efficient assembly of all these ribosomal constituents into ribosomal particles, under various conditions of cell growth a simultaneous and equimolar production of these components is necessary (1,2). Thus ribosome biosynthesis requires a differential and balanced expression of a great number of different genes. To elucidate the molecular mechanisms involved in the coordinate synthesis of ribosomal proteins in yeast several ribosomal protein genes have been isolated by molecular cloning (see Ref. 1 for a review).

From the characterization of DNA fragments carrying ribosomal protein genes it had become apparent that most ribosomal protein genes in yeast are not clustered (3,4,5). In addition, many ribosomal protein genes turned out to be duplicated (3,5).

This paper deals with two ribosomal protein genes, encoding the large subunit protein rp28 and the small subunit protein S16A (rp55), which are exceptional in that they are physically linked. Strikingly, the duplicate

genes are also adjacent to each other suggesting a functional linkage of these ribosomal protein genes. Therefore we decided to investigate the structure and organization of these linked ribosomal protein genes.

Studies of cloned ribosomal protein genes carried out thus far have revealed some common structural features. Most, but not all ribosomal protein genes contain an intron of 300-500 bp near the 5'-end of the coding sequence (3, 5-10). The intron-exon boundaries show conserved sequences similar to the splice sites found in higher eukaryotes. In addition the introns contain near the 3'-end a conserved sequence, TACTAACA, which probably is involved in splicing of the mRNA (11-13).

Computer analysis of the DNA sequences upstream from a number of yeast ribosomal protein genes revealed several homologous sequence elements which may be involved in the regulation and/or expression of these genes (10). As will be demonstrated in this paper the genes coding for rp28 and S16A contain many of the common characteristics of yeast ribosomal protein genes. Apart from that, the duplicated gene pairs show some features which may be unique to these ribosomal protein genes.

MATERIALS AND METHODS

Nomenclature

Yeast ribosomal protein rp55 (14) is identical with S16A in the standard numbering system based on Kaltschmidt and Wittmann 2D-gels (15); rp28 cannot be identified in this gel system. However, rp28 is resolved on the gel system of Gorenstein and Warner (3,16). We have chosen to use the rp28-S16A nomenclature throughout this paper.

Genes isolated from the yeast strains Saccharomyces carlsbergensis and Saccharomyces cerevisiae are referred to as S.ca and S.ce genes, respectively.

Yeast strains

Yeast strains used for the isolation of DNA were Saccharomyces carlsbergensis (NCYC74) and Saccharomyces cerevisiae A364A (ATCC22244).

Isolation of genes

The initial isolation of the first-copy linked genes from S.ce and S.ca have previously been described (3,17). To isolate the second-copy genes for rp28 and S16A of S. carlsbergensis a library was constructed using cosmid pHC79 (Boehringer Mannheim) as a vector. Yeast DNA was isolated as described by Verbeet et al. (18), partially digested with restriction endonuclease Sau3A and size-fractionated by sucrose-gradient centrifugation (19). Appropriate fractions of this yeast DNA were treated with bacterial alkaline phosphatase

and then ligated with a mixture of left-handed and right-handed vector ends (incapable of self-ligation; 20). After packaging in λ phage particles (19) the recombinant DNA was transduced to *Escherichia coli* 1046 (kindly provided by Dr. G.J.B. van Ommen). Colony screening was performed as described previously (20), using a 2.3 kb HindIII-generated fragment isolated from pBMCY44 as a probe. This fragment carries the genes encoding rp28 and S16A and was labelled in vitro by nick translation (21).

Appropriate fragments of the selected recombinant cosmid and of the recombinant bacteriophage A12 which carries the S.ca gene pair (3) were subcloned into pBR322.

Recombinant plasmids were purified from Triton-treated bacterial spheroplasts by CsCl ethidium bromide density gradient centrifugation (22).

Heteroduplex analysis and electron microscopy

Heteroduplex analysis and electron microscopy were performed essentially as described by Verbeet et al. (18).

DNA sequence analysis

DNA sequence analysis was performed using the chain termination method (23). Single-stranded templates were obtained by transforming JM101 or JM103 cells with recombinant bacteriophage M13 RF DNA (24). The M13 vectors used were M13 mp8, mp9, mp10, and mp11 (25).

Restriction site mapping

Restriction endonucleases were obtained from Bethesda Research Laboratories, New England Laboratories or Boehringer (Mannheim) and used as recommended by the supplier. Cleavage sites for AluI, HpaI, MspI (HpaII) and TaqI were determined by the partial digestion procedure of Smith and Birnstiel (26).

Southern blot analysis

Total yeast DNA was isolated from both *S. carlsbergensis* and *S. cerevisiae* and digested with EcoRI or HindIII. The resulting fragments were electrophoresed on 1% agarose gels, transferred onto nitrocellulose (27) and hybridized with 32 P-labelled DNA fragments as described previously (6).

RESULTS AND DISCUSSION

Southern analysis

The linked genes coding for ribosomal proteins rp28 and S16A have been isolated from a colony bank of HindIII-generated S.ca DNA fragments in pBR322 as described previously (17). The genes have been mapped on plasmid pBMCY44 by electron microscopic R-loop analysis (17) as shown in Fig. 1A. To determine the copy number of the genes coding for the two ribosomal proteins, genomic

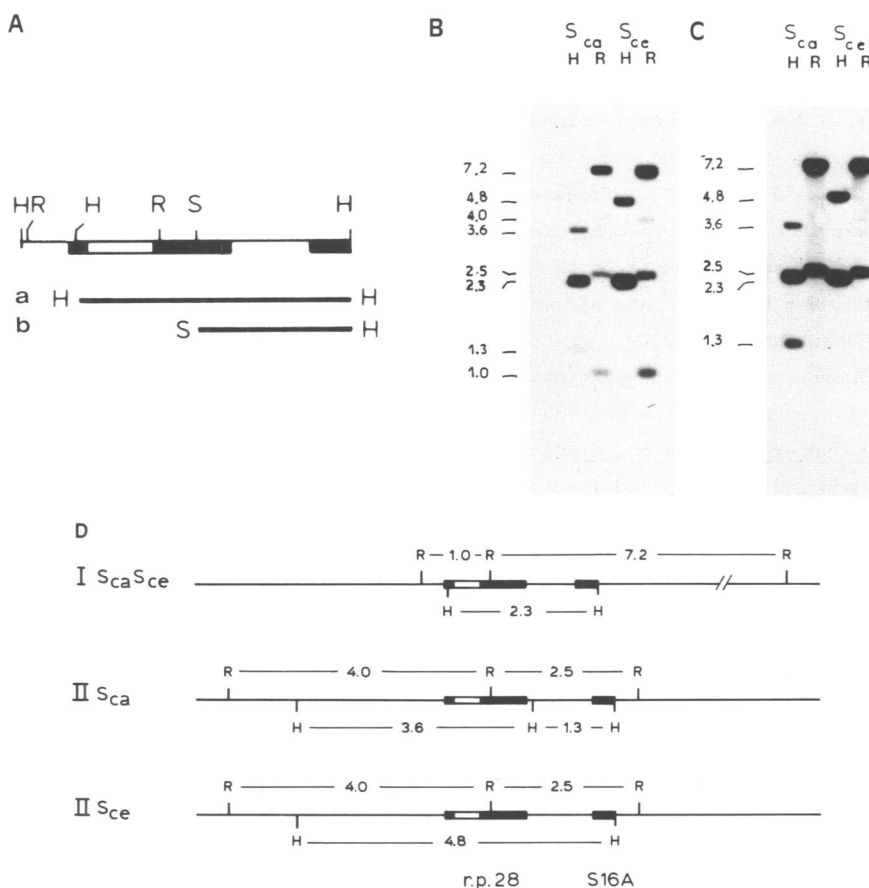


Fig. 1. Copy number analysis of the genes encoding rp28 and S16A.

In A the location of the *S.ca* *rp28* and *S16A* genes on the physical map of the insert of recombinant pBMCY44 is depicted (17). Symbols for restriction sites are H (HindIII), R (EcoRI) and S (SacI). The intron in the *rp28* gene is indicated by an open bar. The results of the hybridization of 32 P-labelled fragments a and b (panel A) to *S.ca* and *S.ce* DNA digested with HindIII (H) or EcoRI (R) are shown in panel B and C, respectively. I *S.ca* is the region in the genome corresponding with the insert of pBMCY44 as well as A12 in panel D. II *S.ca* and II *S.ce* are the regions of the genome corresponding to duplicate copies of *rp28* and *S16A* in the two strains based on Southern analysis and EM R-loop analysis. Note that the 5' small exon of *S16A* cannot be detected by R-loop analysis but can be demonstrated by heteroduplex and sequence analysis (Fig. 3 and 4).

Southern analysis was performed using a 2.3 kb HindIII-generated fragment as well as a 1.3 kb (SacI plus HindIII)-generated fragment from pBMCY44 as a probe. The autoradiograms showing the result of hybridization to genomic digests of both *S.ca* and *S.ce* DNA are presented in Figs. 1B and 1C. It can be

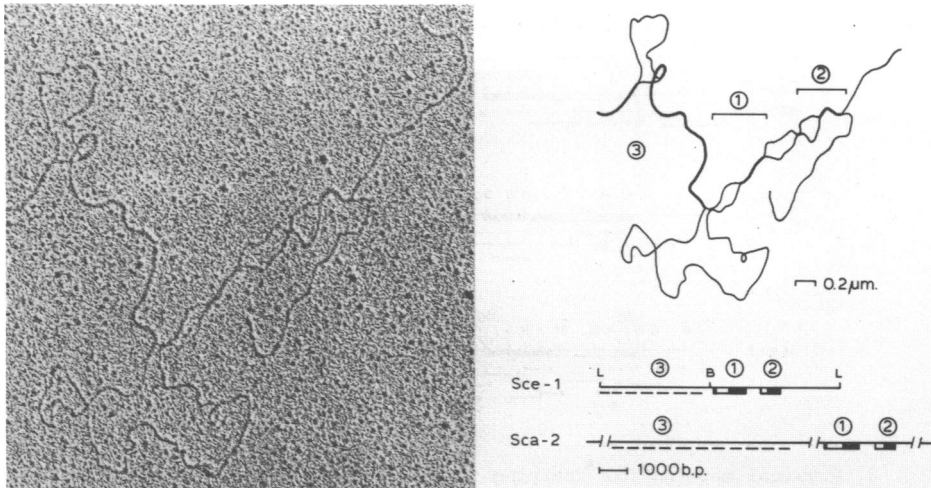


Fig. 2. Heteroduplex analysis of duplicate genes encoding rp28 and S16A. Recombinants *S.ce*-1 and *S.ca*-2 carry duplicate gene pairs coding for rp28 and S16A. *S.ce*-1 was linearized by cleavage with SalI; the 40 kb recombinant cosmid *S.ca*-2 was linearized by random shear. Only amino acid-coding regions of the rp28 (1) or S16A (2) gene and regions of homologous vector DNA (3; pBR322 and pHC79) form stable duplex structures. Similar results were obtained in the comparison of the two copies of the gene pair isolated from *S. carlsbergensis*.

concluded that portion of both genes is duplicated in the genome of both yeast strains and, in addition, that most probably the genes are also adjacent at the second locus. This agrees with the published results for *S.ce* (3). The latter finding suggests a functional linkage of the genes coding for S16A and rp28. The interpretation of the hybridization results described above is illustrated in Fig. 1D.

The duplicate gene pair was isolated from a cosmid bank as described in the Materials and Methods section. Isolation of the gene pair from *S. cerevisiae* that corresponds with the insert of pBMCY44 has been described previously (3). The *S. cerevisiae* clone carrying copy 1 of rp28 and S16A is designated A12.

We performed a comparative analysis of the structure and organization of the genes coding for rp28 and S16A to get insight into the pattern of conserved and diverged sequences in both the duplicate gene pairs and the two different yeast strains.

Heteroduplex analysis

First heteroduplex analysis under the electron microscope was performed. On

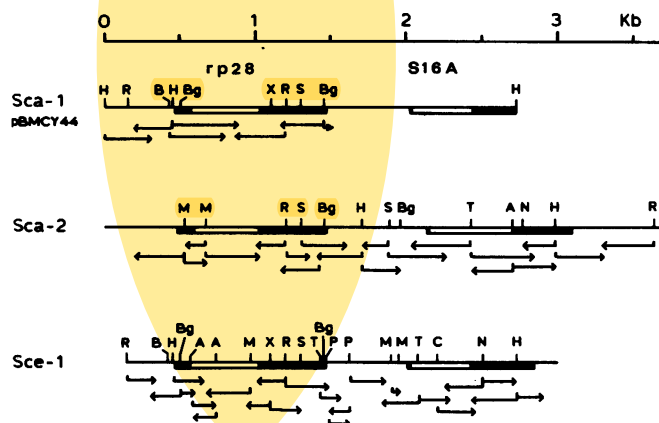


Fig. 3. Physical maps of DNA carrying the rp28 and S16A genes and the DNA sequencing strategy.

Symbols for restriction endonucleases are A (AluI), B (BamHI), Bg (BglII), C (BclI), H (HindIII), M (MspI, HpaII), N (HincII), P (HpaI), R (EcoRI), S (SacI), T (TaqI) and X (XbaI). Introns and exons are indicated by open and solid bars, respectively.

S.ca-1 Map of the yeast DNA insert of pBMCY44 (17).

S.ce-1 Map of part of the insert of the recombinant phage A12 (3).

S.ca-2 Map of part of the recombinant cosmid carrying the second-copy genes for rp28 and S16A (this paper).

The arrows indicate the direction and extent of nucleotide sequence analysis.

this level of examination the corresponding DNA fragments from S.ce and S.ca turned out to be completely homologous (results not shown). Heteroduplex analysis of the duplicate gene pairs, however, indicated that, apart from the presumed coding regions, they have very little sequence homology (see Fig. 2). The electron micrograph suggests that both genes are interrupted by an intron (as has been established previously for the rp28 gene [17]). Unlike the exons, which are highly conserved and therefore form stable heteroduplexes, the intervening and flanking sequences including the intergenic region show a high degree of divergence since no stable duplex structures of these regions can be observed. Since it is a general characteristic of yeast ribosomal protein genes studied so far that these genes are split by an intron near the 5'-side of the coding sequence, the heteroduplex analysis predicts a head to tail arrangement of these genes.

Sequence comparison: S.ca vs S.ce

The respective restriction maps of the cloned recombinant DNAs are presented in Fig. 3. Correlation of the physical maps with the results of the genomic Southern analysis (cf. Fig. 1D) confirmed our preliminary conclusion with

[illegible]

1	GCT GGT GCC AGA GCC AAG ATT GTT AAG GCT GGT GGT GAA TGT ATC ACT TTG GAT CAA TTA GCT GTC AGA GCT CCA	1162
2	*** **	1162
3	GCT GGT GCC AGA GCC AAG ATT GTT AAG GCT GGT GGT GAA TGT ATC ACT TTG GAT CAA TTA GCT GTC AGA GCT CCA	1126
	<i>ala gly ala arg ala lys ile val lys ala gly gly glu cys ile thr leu asp gln leu ala val arg ala pro</i>	
	120	
1	AAG GGT CAA AAC ACT TTG ATC TTG AGA GGT CCA AGA AAC TCC AGA GAA GCT GTC AGA CAC TTC GGT ATG GGT CCA	1237
2	*** **	1237
3	AAG GGT CAA AAC ACT TTG ATC TTG AGA GGT CCA AGA AAC TCC AGA GAA GCG GTC AGA CAC TTC GGT ATG GGT CCA	1201
	<i>lys gly gln asn thr leu ile leu arg gly pro arg asn ser arg glu ala val arg his phe gly met gly pro</i>	
	140	
1	CAC AAG GGT AAG GCT CCA AGA ATC TTG TCC ACC GGT AGA AAG TTC GAA AGA GCT AGA GGT AGA AGA AGA TCT AAG	1312
2	*** **	1312
3	CAC AAG GGT AAG GCT CCA AGA ATC TTG TCC ACC GGT AGA AAG TTC GAA AGA GCT AGA GGT AGA AGA AGA TCT AAG	1276
	<i>his lys gly lys ala pro arg ile leu ser thr gly arg lys phe glu arg ala arg gly arg arg ser lys</i>	
	160	180
1	GGT TTC AAG GTG TAA GTTAACTGA- AATGAAAA-- ----- TTTCATATTT ACTTTTTTAT TGTACTCAT TTGTAATTCA	1384
2	*** **	1384
3	GGT TTC AAG GTG TAA TCTAGTATGG TTTGAAACCT TACAATTTTT CTTCCTTGT CTCTTTTCT TGTTCAGTG TATATTAGGT	1361
	<i>gly phe lys val end</i>	
	186	
	+100	
2	TAAACTACAT ACACCTTCAA TCGTTTCTTT CAAACTACAT AATTTTTCTT GCGCATCAAT AACGCAITTA GTTCATAAAG TGAGTCAAAG	1474
3	TGGGAA----- -AGA GGGATTTTTC CATACCATAT GACTGACTAC AATATATACA TGTATAATAA CTTCATAATC TAAACCAACC	1440
	+100	
	+200	
2	TTAACACTAG AATATTGCT ACACCATCAA TAGGCTAGAC CATAGTTGAA AACTTNCATA ATAAATTCCT CCGTTTTCAA TC-TTCATAT	1563
3	TATCAGTICA GTATCAAG-- ----- TCAACTATTC CGCCCTATGC ATAAACCTAC TAAACTATCA TTCTTCACAC	1508
	+200	
	-300	
2	ACTGTGCTC TA---ACCAT GATACCGTGA CACAAC TACA TCCGTACACA TGTGACGTTT GTTCAACCCG TACATTTATA TAAACCGTT	1650
3	TTCACAAAGC TTTTCCCAIT TTTTTCATA CTACTTTACA TCCGAACATT TTAGAAACCC ACACCAT--- ---ATACCTT	1582
	-400	
	-200	
2	CTGGCGCCT TTTATTTTTT TACATTCTCT ATGATCGGGA TTGCAGAAG CCGTG----- -----	1705
3	TGTTGCACTA TTGATTTCCT TCCTGATGTC AGCTTTTTGT GCTTTGACAA AAAAATCGCG TCTACGTCGG TCCGTTCTCC CTGAATCAAT	1672
	-300	
2	----- --AAATTTTT CAATGTGAGG TTCGCCCT-- --TGTTTCG AAAAGCCCTA	1748
3	TAGCGCGGTT TGAGCCACG AGGACGGAGC TCTAGTGACA AGCCCTGGTG TTTGGTGAGG TAATGCACAT TCGTGTCTCT TTCTACTGTA	1762
	-200	
	-100	
2	TTGAGATACC GGAAAGATAT AGGTGAAATG AAGAAAA--C TATGGTGTGT ATATCTAATA CCCCGTTCGG TTTATTAATA TTTTAGCTTG	1836
3	TTGAGATCTC CAGTTTACGG CTCCTGGGT GCCACCCGTA ACCTGGTTGG ----- -TGTC CCCATTTCAA	1826
2	AAAGCGAAG- ----- -TGATACGAT	1854
3	TAAAGCAACA TTAGTGAAGA CACAATCGTT AAAATGGACT AATGAAATTT TAAAGTGCGA TTTTGTGAA TATTGACAC AAAGGTATAG	1916
	-100	
2	CGACAAATAG AGTAAAA ATG CCA GGT GTT TCC GTT AG GTACGT AAAAAATGACA TATCAT--AA TGATTATTAC TAT-ATGTTT	1934
3	AACCAAGAT AATAAAG ATG GCA GGT GTT TCC GTT AG GTACGT ATAACCTTCA CT-CATTGAA TGAGTATTIA GAGAATGAAT	1998
	<i>met pro gly val ser val arg</i>	
	1 ala 7	
2	CGAGTGGTTA GATGGAAATA AAACGCACTC ATTCCAGCAG CATAGACACG CAAGCAAAGT ACTTCTTC-- -----	2002
3	GGA--TATTA GGTGGAAAC AACGAGATGA AACATATAG GATTGAGAAA AAGGAATTAG TGCAGAATTT TCCATTACC TGATAAATTG	2086
2	----- --GTCAAGTT AAATAGCACC CATGGTTAAA TCAATGATAT ATCAACTGTC CCATA-ATAA	2059
3	AAGTTCATCT TTACTGTGCA TTTACCGAAA CGAATGAGTT AAACATTTAT GATATAATCG CTGTAATGT GGAGAGAATA TTTTGAGCCA	2176
2	TTGAAACAAA ATATTGATCA AATTTCGCGA ATGAGTGCAA TGTGTAAATA TATGGAAGAA GAGGGAGATT AAGGGTAGTG GTGCTATTGT	2149
3	ACTATACAAA ATGAAG----- -TT CTCAGACGAA GAGGTAATCA TTTATTATT GGAATCGAAT	2234

2	CTCTT-----	-----TGTAT	TACTAGGATT	TCAATTCCTT	CCGATAAGTT	CTCATGAAAC	-----	2200
3	GATAGGAGGT	TTTAACTAGG	TGGGATGTTT	TCTAATGAAA	TAGGAGAACC	TGAACCAAGA	CGTTTAAAGT	2324
							GGAAGGAAAT	TTGTAACCGT
2	-GTAATG--	-----	-----GTTAAT	GAAAG-----	-----	-----	-----	2217
3	GGTAATGCAA	AGCAATGATA	AGCGATTTTA	TGCCGTTAAT	TTAAGCAACA	ACCCGAGACC	ACGATACTGC	2414
							CGCACCAAGA	AGTTGTCTCA
2	-----	-----TAT	TTTTTTTAAA	CTGTATCAIT	TACTAACAAT	AACTTTTTTG	TTTTGATTTT	2279
3	ATAAATATTT	CACTGAGCAT	TCCATATTCT	ATTTTTACGA	GAAAACCTTT	TACTAACAAA	AGCATTTCAA	2502
							TTT--CAATT	
2	AG	A GAC GTT GCT GCT CAA GAT TTC ATT AAT GCT TAC GCT TCT TTC TTG CAA AGA CAA GGT AAG CTA GAA GTT	2351					
3	AG	A GAC GTT GCA GCT CAA GAT TTC ATC AAT GCT TAC GCT TCT TTC TTG CAA AGA CAA GGT AAA ITA GAA GTC	2574					
		asp val ala ala gln asp phe ile asn ala tyr ala ser phe leu gln arg gln gly lys leu glu val						
		8 20						
2	CCA GGT TAC GTT GAC ATT GTC AAG ACC TCT TCT GGT AAC GAA ATG CCA CCA CAA GAT GCC GAA GGT TGG TTC TAC	2426						
3	CCA GGT TAC GTT GAC ATT GTC AAG ACC TCT TCT GGT AAT GAA ATG CCA CCA CAA GAT GCC GAA GGT TGG TTC TAC	2649						
		pro gly tyr val asp ile val lys thr ser ser gly asn glu met pro pro gln asp ala glu gly trp phe tyr						
		40						
2	AAG CGT GCT GCC TCT GTT GCC AGA CAC ATT TAC ATG AGA AAA CAA GTT GGT GTT GGT AAA TTG AAC AAA TTA TAC	2501						
3	AAG CGT GCT GCC TCT GTT GCC AGA CAC ATT TAC ATG AGA AAA CAA GTT GGT GTT GGT AAA TTG AAC AAA TTG TAC	2724						
		lys arg ala ala ser val ala arg his ile tyr met arg lys gln val gly val gly lys leu asn lys leu tyr						
		60 80						
2	GGT GGT GCC AAG AGC AGA GGT GTT AGA CCA TAC AAG CAC ATT GAC GCT TCC GGT TCT ATC AAC AGA AAG GTT TTG	2576						
3	GGT GGT GCC AAG AGC AGA GGT GTT AGA CCA TAC AAG CAC ATT GAT GCT TCC GGT TCT ATC AAC AGA AAG GTT TTG	2799						
		gly gly ala lys ser arg gly val arg pro tyr lys his ile asp ala ser gly ser ile asn arg lys val leu						
		100						
2	CAA GCT TTG GAA AAG ATT GGT ATC GTC GAA ATC TCT CCA AAG GGT GGT AGA AGA ATC TCT GAA AAC GGT CAA AGA	2651						
3	CAA GCT TTG GAA AAA ATT GGT ATT GTC GAA ATC TCT CCA AAG GGT GGT AGA AGA ATC TCT GAA AAC GGT CAA AGA	2874						
		gln ala leu glu lys ile gly ile val glu ile ser pro lys gly gly arg arg ile ser glu asn gly gln arg						
		120						
2	GAT TTG GAT CGT ATT GCC GCT CAA ACT TTG GAA GAA GAC GAA TAA ATAT AAAATCTATA ATTTATATAT ATATACTACT	2730						
3	GAT TTG GAT CGT ATT GCC GCT CAA ACT TTG GAA GAA GAC GAA TAA GCGA GTTGTTGTTAAA ATAAAGCT A-AGATTTTTT	2952						
		asp leu asp arg ile ala ala gln thr leu glu glu asp glu end						
		140 144						
2	ACTACAATTA TCATCATACA AGTAATAATA A-----	2761						
3	ATTAATATTA TCATTATACT ATAGTATTAT CGTTTAACTC AAACAAAATT GATTTTAAAT AATTTATTTA TTAAATTTT GCCATTTTAT	3042						
		+100						
		+200						
3	ATGTACATTC ATCATCATCA ATATAGCCGA AACCAAATA AATAGATTTT GTTTTTTTAA GATAGAATGT GTCAAACCAT GGATTGCGAT	3132						
		+300						
3	CTGGATAAAA ATGACAACCTT TTAAGTTTTC GTCTACAAAA CATTAAAAAC ACCATTGAAG AATAAATGGT AGTAAATAAT TTTACAATGA	3222						

Fig. 4. Primary structure of the *rp28* and *S16A* gene pairs of two yeast strains.

1, 2 and 3 represent the primary structures of *S.ca-1*, *S.ce-1* and *S.ca-2* (cf. Fig. 3), respectively. The nucleotides for *S.ca-1* and *S.ce-1* are indicated separately only in the case of base substitutions. Dashed lines indicate deletions. The positions at which both copy genes (*S.ce-1* and *S.ca-2*) have identical nucleotide sequences are marked by asterisks. Notable sequences are referred to in the text.

||||| TATA-like elements (36).

||||| PyAAPu (37,38)

~~~~~ putative transcription-termination and/or polyadenylation signals (10,39,41).

respect to the duplicate and linked nature of these genes. Southern and heteroduplex analyses permitted us to map the genes as indicated in Fig. 3. To analyse the structure of the genes coding for rp28 and S16A the complete nucleotide sequences of the S.ce gene pair and the second-copy S.ca gene pair have been determined. To be able to evaluate a comparison of duplicate gene copies isolated from two yeast strains, we also have sequenced the "first-copy" S.ca rp28 gene carried by pBMCY44 that corresponds with the S.ce rp28 gene. In Fig. 3 the strategy to determine the complete sequence is outlined. The primary structure of the rp28-S16A gene pairs that resulted from these sequence analyses is presented in Fig. 4.

First we compared the sequence of the homologous S.ca and S.ce gene copies coding for rp28. Only 13 nucleotide differences were observed in the non-translated and flanking regions of the gene and none in the coding region. Three of these changes were in the intron and ten were in the 300 nucleotides 5' to the coding region (Fig. 4). A low degree of divergence on the DNA level between S.ca and S.ce has been reported previously for the actin gene (28) and the ribosomal RNA genes (29) and was also observed for the histon H4 gene (30,31). Our results support the present idea that S.ca and S.ce are similar or identical yeast strains (32). Therefore any alterations (except for minor base substitutions) found by comparing the DNA sequences of the first-copy gene pair of S.ce with the second-copy gene pair of S.ca can be considered as differences between the duplicated gene pairs rather than strain differences.

The sequence data presented in Fig. 4 indicate that the genes coding for rp28 and S16A are transcribed in the same direction in both gene pairs and that there are 544 and 642 base pairs between coding regions in S.ce gene pair 1 and S.ca gene pair 2, respectively.

### Sequence comparison: duplicate genes, coding region and intron

The genes contain some structural features which many ribosomal protein genes have in common. First of all, as predicted above, both genes are interrupted by an intron near the 5'-end of the coding sequence. These data are summarized in Table I. The intron-exon boundaries are conserved, also near the 3'-end of the introns another common sequence, TACTAACA, is present. This sequence has been shown to be essential for splicing of intron-containing pre-mRNAs in yeast (11-13). It is clear that the intervening sequences interrupting the duplicate genes differ not only in length (cf. Table I) but also in nucleotide sequence. By computer analysis some regions could be identified that are conserved only in the introns of duplicate genes encoding the same protein. Two such conserved regions within the rp28-introns show a dyad symmetry of

Table I. Structure of the ribosomal protein genes encoding rp28 and S16A.

|            | 5'                |                                              | 3'               |
|------------|-------------------|----------------------------------------------|------------------|
| Sce rp28:  | ATG - 36 codons - | ↓ GTATGT - 411 n. - TACTAACA - 19 n. - TAG ↓ | 149 codons - TAA |
| Sca rp28:  | ATG - 36 codons - | GTATGT - 377 n. - TACTAACA - 35 n. - CAG     | 149 codons - TAA |
| Sce S16A:  | ATG - 5 codons -  | GTACGT - 343 n. - TACTAACA - 30 n. - CAG     | 138 codons - TAA |
| Sca S16A:  | ATG - 5 codons -  | GTACGT - 505 n. - TACTAACA - 29 n. - TAG     | 138 codons - TAA |
| consensus: | GTAPyGT           | TACTAACA                                     | PyAG             |

13 base pairs, which would permit a secondary structure element in the precursor rp28-mRNAs (indicated in Fig. 4 with arrows).

The introns interrupt the coding regions of the duplicate genes at the same position (*cf.* Table I).

With respect to the amino acid coding regions, we observed 95% homology between both duplicate gene pairs. Most base substitutions do not give rise to an amino acid substitution. For the S16A genes only 1 out of 21 nucleotide substitutions causes an amino acid substitution, *viz.* Pro/Ala at position 2 (*cf.* Fig. 4). Concerning the rp28 genes, the 27 base changes do not result in amino acid substitutions. It is interesting to know that the silent base changes in rp28 are clustered in the short 5'-exon. 14 out of 112 bases are substituted in the first exon and only 13 out of 446 are changed in the large exon. This clustering of silent base changes is not seen in S16A. By Otaka *et al.* (33) the amino acid sequence of the first 49 N-terminal residues of the S16A protein has been established. Their data are completely consistent with the amino acid sequence deduced from the DNA sequence. Moreover, Otaka *et al.* found both proline and alanine as N-terminal amino acids at a ratio of 1:2. This finding is in excellent agreement with our results and, moreover, provides evidence that both S16A genes are expressed.

The coding sequences of all four genes examined show a rather preferred codon usage consistent with previous findings for other efficiently transcribed yeast genes (34,7).

#### Sequence comparison: noncoding regions

Comparison of the regions upstream from the ATG initiation codons only reveal short regions that are conserved between duplicate genes encoding the same protein. The presumed leader sequences of the duplicate genes show a low degree of homology as can be deduced from Fig. 4. It is unlikely, therefore, that specific primary structure elements in this part of the mRNAs are involved in translational control in the autogenous way it has been observed in bacterial cells (35).

In the 5'-flanking region of the S16A genes the most remarkable conserved

sequence is TACATCCG(T/A)ACA at positions -274 and -389 in gene copy 1 and gene copy 2, respectively (Fig. 4). This sequence is very similar to consensus sequence HOMOL1 observed by Teem *et al.*, *viz.* AACATC(T/C) (G/A)T(A/G)CA (10). Since this sequence element occurs at a position of about -300 for 6 yeast ribosomal protein genes investigated so far and does not precede any of the 20 yeast non-ribosomal protein genes examined, it was proposed to play a role in the coordinate expression of these genes (10). A similar sequence, GACCTCAGTACA, matching in 9 out of 12 positions with consensus sequence HOMOL1 was observed at -197 from the rp28-2 coding region (Fig. 4), but not in the 5' flanking region of the rp28-1 gene. In the upstream region of both rp28 genes, however, we observed the conserved sequence T(T/C)TTTCTTGCTGGAG (A/C)TT at position -193 and -226 in the first- and second-copy genes, respectively (Fig. 4). We suggest that additional sequence elements of this kind may function in the coordinate expression in subsets of yeast ribosomal protein genes.

Sequences that fit other consensus sequences proposed by Teem *et al.* (10) are AATTTTCA at position -165 in the S16A-1 gene (consensus sequence HOMOL4: (T/A)AT(T/A)TTnCA), TATTTA at position -97 in rp28-2 and TATT(T/A)(T/A) at positions -47 and -53 in the S16A-1 gene (consensus sequence HOMOL5: TATT(T/A)(T/A), resembling the "TATA" box [36]). More general TATA-like structures are found in the two duplicate pairs of genes that lack consensus sequence HOMOL5 (Fig. 4). Finally, in each gene one or more sequences can be identified that fit the consensus sequence PyAAPu that has been proposed to act as a transcriptional start site for yeast polymerase B (37,38) (see Fig. 4).

From the sequence data presented in Fig. 4 it is apparent that the sequences downstream of the TAA stopcodon have diverged to a similar extent as the other noncoding regions. Several sequences have been proposed to be involved in transcription termination and/or polyadenylation in yeast (10,39,40) or *Eumetazoa* (41). All four genes examined contain one or more of these consensus sequences as indicated in Fig. 4.

In summary, we have analysed the structure of two linked ribosomal protein genes in yeast by heteroduplex - and sequence analysis. We have demonstrated that these genes are duplicated in the genome in the same linkage arrangement. The almost complete conservation of amino acid sequence in duplicate copies of these genes, as deduced from the DNA sequence, and the work of Otaka *et al.* (33) described in the results section, strongly suggests that all four genes make functional products. The fact that a similar linkage arrangement is

maintained in both gene pairs, even though the distance and sequence between them is different, suggests that the linkage arrangement may play some role in the regulation of expression of these genes. Finally, it will be of interest to determine whether these duplicated genes, which code for almost identical gene products, are regulated in the same or different manner, considering that the regions 5' to the ATG start codons are almost totally diverged.

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